

# Inheritance and molecular mapping of leaf rust resistance gene in hexaploid wheat Synthetic 45

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#### Abstract

To contain the infection of newly evolved leaf rust virulence(s), diverse source of germplasm need to be explored for identification of novel genes. In order to exploit wild resources, CIMMYT has developed a series of synthetic hexaploid wheat (SHW) by combining the accessions of D genome donor Triticum tauschii and T. durum. In the present study, Synthetic 45, was evaluated for effectiveness of its resistance against diverse leaf rust pathotypes. Mode of inheritance of resistance to understand the nature of genetics and molecular mapping to locate its chromosomal position were studied. Characterization of leaf rust resistance in Synthetic 45 by multi-pathotype tests showed a high degree of seedling resistance to 20 diverse pathotypes of leaf rust pathogen and adult plant resistance against two most prevalent pathotypes, 77-5 and 104-2. Inheritance studies showed that resistance in Synthetic 45 was governed by a single recessive gene. Molecular mapping and linkage with microsatellite markers, Xwmc432 and Xcfd15 have indicated that the resistance gene is located on short arm of 1D chromosome with 6.1 cM distal to Xwmc432 and 10.6 cM to Xcfd15 with 4.6 cM distance among the two markers. The gene identified in Synthetic 45 has been tentatively designated as LrSyn45.

Key words: Synthetic Hexaploid Wheat, leaf rust resistance, inheritance, mapping, wheat rust

# Introduction

Three wheat rusts (leaf, stem and stripe rusts) continue to pose a serious threat by inflicting yield losses in different parts of world. Among the three, leaf rust caused by *Puccinia triticina* Eriks. is most common and widespread on wheat (*Triticum aestivum* L.) in India (Tomar et al. 2014). Depending upon the severity and duration of infection, the losses may reach up to 50% of the yield. In India, 51 pathotypes of leaf rust have been reported during 1931-2016, out of which 15 new pathotypes have been identified in nearly last 10 years. Many useful resistance genes like Lr9, Lr19 and Lr28 have been rendered ineffective (Nayar et al. 2003; Bhardwaj et al. 2005, 2010a and 2011). Pathotype 77-5 (121R63-1) is currently the most virulent and frequent in the Indian subcontinent, which knocks down several leaf rust resistance genes like, Lr1, Lr2a, Lr2b, Lr2c, Lr3a, Lr10, Lr11, Lr14a, Lr14b, Lr15, Lr16, Lr17a, Lr20, Lr23, Lr26, Lr27+Lr31, Lr33, Lr36, Lr38, Lr43 and Lr44 (Bhardwaj et al. 2010b). Further, most of the native T. aestivum leaf rust resistance genes became ineffective in India to the most virulent and prevalent pathotypes of group 77. Only a few leaf rust resistance genes viz., Lr1, Lr3, Lr9, Lr10, Lr13, Lr14a, Lr17, Lr19, Lr23, Lr24, Lr26, Lr28 and Lr34 have been exploited in the leading Indian wheat varieties (Bhardwaj et al. 2010c). Among the major seedling resistance genes utilized, only Lr24 is having no known virulence in India, however, virulence is known in other parts of the world (Singh 1991).

In a scenario of rapid virulence development against major genes, searching new sources of resistance particularly from related species of wheat shows immense potential. Goat grass (*Triticum tauschii*), the D-genome donor to bread wheat, a relatively untapped germplasm pool was used for production of synthetic hexaploid wheats (SHWs) at CIMMYT. The SHWs possess potential variability for morpho-agronomic traits as well as resistance/ tolerance to biotic/abiotic stresses (Valkoun et al. 1990; Cox et al. 1994). The Division of Genetics, IARI,

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New Delhi maintains an elite set of 65 SHWs. One of these SHWs, designated as Synthetic 45 was observed to be resistant to leaf rust in field conditions under artificial epiphytotics with most virulent and prevalent pathotype, 77-5(121R63-1) of leaf rust. Therefore, the present investigation was undertaken to study effectiveness of resistance in Synthetic 45 against an array of leaf rust pathotypes, to determine nature and number of gene(s) controlling resistance and to locate the position of resistance gene(s) by linked microsatellite markers.

#### Materials and methods

# Development of populations for inheritance and mapping

Leaf rust resistant Synthetic 45 (68.111/RGB-U// WARD/3/FGO/4/RABI/5/Ae. squarrosa), was evaluated against diverse pathotypes of leaf rust to identify resistance gene(s) and its mapping on a specific chromosome. Highly susceptible genotypes, Agra Local and Kharchia Local were used as contrasting parents for development of segregating populations involving Synthetic 45. Pure inoculum of 20 pathotypes of leaf rust pathogen *P. triticina* Eriks. used in the study for multipathotype testing was obtained from IIWBR, Regional Station, Flowerdale, Shimla. The most virulent pathotype, 77-5 was selected for inheritance studies.

Resistant parent Synthetic 45 was crossed with susceptible parents, Agra Local and Kharchia Local during *rabi* 2014-15 at IARI, New Delhi. The  $F_1$  plants were raised during summer 2015 at IARI Regional Station, Wellington, The Nilgiris, Tamil Nadu to obtain  $F_2$  seeds for genetic analysis. The  $F_1$  hybrids were backcrossed to susceptible parents to obtain BC<sub>1</sub>F<sub>1</sub> generations in off-season (summer nursery) at Dalang maidan, Lahaul Spiti, Himachal Pradesh. The  $F_1$ ,  $F_2$ and BC<sub>1</sub>F<sub>1</sub> generations were raised and tested along with parents at seedling stage under glass house conditions at IARI, New Delhi during *rabi* 2015-16.

# Multipathotype testing and screening of populations against leaf rust

For multipathotype testing, seedlings of resistant parent Synthetic 45, susceptible parents Agra Local and Kharchia Local were raised in 10 cm pots, while for inheritance studies  $P_1$ ,  $P_2$ ,  $F_1$ ,  $F_2$  and  $BC_1F_1$ seedlings were raised in the rectangular trays (28cm ×10cm× 7.5cm). Ten days old seedlings were inoculated with 20 pathotypes separately by spray as suspension of rust uredospores in water along with a drop of Tween 20 (Polysorbate 20). Inoculated pots were kept in humidity chamber for 48 hours before shifting to glass house benches (Joshi et al. 1988). Infection types were recorded after 12 days of inoculation following 0-4 scale classification as per Stakman et al. (1962). The infection type 0, ;, 1 and 2 were classified as resistant reaction while infection types 3 and 4 were classified as susceptible.

#### Genetic analysis

For inheritance studies, the  $F_1$ ,  $F_2$  and  $BC_1F_1$ generations along with parental lines were tested at seedling stage against pathotype 77-5 of leaf rust pathogen. Chi-square test was applied to observed  $F_2$ phenotypes to determine goodness of fit for a ratio according to Panse and Sukhatme (1967) and to compare the observed ratio with that expected for Mendelian segregation for leaf rust resistance gene. To verify the  $F_2$  results and proposed hypothesis  $BC_1F_1$  generation was genetically analysed.

#### Genomic DNA isolation and PCR amplification

All the tested F<sub>2</sub> seedlings of the cross Synthetic 45 × Agra Local were transplanted in field for molecular mapping. Tender and fresh leaves were collected from 40-45 days old plants and DNA was isolated following CTAB method (Murray and Thompson, 1980). The RNA was digested and removed by treating the samples with DNase free RNase (10 mg/ml) denatured at  $70^{\circ}$ C. The purified DNA was quantified on 0.8% agarose gel by loading 1µl of DNA samples, along with known quantity of  $\lambda$  uncut DNA (100ng, 200 ng). The quantified DNA was diluted with TE buffer to a working concentration of approximately 25ng/µl. The PCR for SSR marker analyses were performed with 10  $\mu$ l reaction volume in the 96-well PCR plates with thermal seal in thermal cycler (model Master cycler pro S, Eppendorf) at temperature profile of 94<sup>0</sup>C for 4 min for initial denaturation, followed by 45 cycles having Ist step at 94°C for 1 min, II<sup>nd</sup> step at specific annealing temperature for 1 min and III<sup>rd</sup> step at 72°C for 1 min, with a final extension after 45<sup>th</sup> cycle at 72°C for 10 min. The amplified PCR products were subsequently resolved on 3.5% metaphor gel in 1X TBE buffer and visualized under UV trans-illuminator in a gel documentation system (Gel Documentation System, Syngene).

# Parental polymorphism and Bulked Segregant Analysis (BSA)

The parental polymorphism survey was carried out between Synthetic 45 and Agra Local using 807 SSR markers covering all the 21 chromosomes of the three (A, B and D) genomes of wheat. The polymorphic markers identified by parental polymorphism were used in BSA as described by Michelmore et al. (1991) to identify putatively linked SSR markers. The resistant and susceptible bulks were constituted by pooling equal amount of DNA from 10 resistant and 10 susceptible  $F_2$  plants, respectively.

#### Linkage analysis and map construction

The markers distinguishing two bulks and parents were considered as putatively linked to leaf rust resistance gene in Synthetic 45. Such putative markers identified in BSA were used for genotyping of  $F_2$  population and linkage analysis was done using MAPMAKER version 3.0 to construct linkage map (Lander et al. 1987) with a minimal LOD score of 3.0 and a maximal genetic distance of 30.0 cM. The genetic distance was calculated in cM using the mapping function of Kosambi (1944).

## **Results and discussion**

# *Effectiveness of leaf rust resistance in Synthetic* 45

At seedling stage Synthetic 45 expressed high levels of resistance (Fig. 1) against all the 20 pathotypes with infection type (IT) ranging from ';' to '1' (Table 1). In contrast, Agra Local and Kharchia Local were susceptible exhibiting ITs '33<sup>+</sup>' to '3<sup>+'</sup> against all the 20 pathotypes considered in the study. The host-pathogen interaction between Synthetic 45 and 20 pathotypes revealed that leaf rust resistance of Svnthetic 45 is broad spectrum having effectiveness against diverse pathotypes of leaf rust pathogen (P. triticina), including the two most predominant pathotypes viz., 77-5 and 104-2 in the natural pathogen population in India. Analysis based on avirulence/virulence formula of 20 pathotypes used in the study and actual infection types produced on Synthetic 45 by host pathogen interaction, indicated that the leaf rust resistance in Synthetic 45 could be a new source of resistance or one among the designated Lr gene(s) not present in the differential set commonly used in to identify new pathotypes of leaf rust pathogen.

At adult plant stage also Synthetic 45 exhibited highly resistant response of 5R (5% severity of rust



Fig. 1. Disease reaction of Synthetic45 against leaf rust pathogen

Table 1.Infection types on Synthetic 45, Agra Local and<br/>Kharchia Local against 20 pathotypes of<br/>Puccinia triticina when tested at seedling stage<br/>of plant growth at mean temperature range 20-<br/>28°C

S.No.	Path desig	notype gnation	Synthetic 45	Agra local	Kharchia Iocal
	Old	New			
1	12A	5R13	;1-	3+	3+
2	12-1	5R37	;1í	$33^+$	33 <sup>+</sup>
3	12-7	93R45	;1-	33 <sup>+</sup>	3+
4	12-8	49R45	;1=	3+	33 <sup>+</sup>
5	12-9	93R37	;1=	3+	33 <sup>+</sup>
6	77-1	109R63	;1-	$33^+$	33 <sup>+</sup>
7	77-2	109R31-1	;1=	3+	3+
8	77-5	121R63-1	;1-	3+	3+
9	77-8	253R31	;1-	3+	33 <sup>+</sup>
10	77-10	377R60-1	1	$33^+$	33 <sup>+</sup>
11	104-1	21R31-1	;1=	3+	3+
12	104-2	21R55	;1 <sup>=N</sup>	3+	33 <sup>+</sup>
13	104-3	21R63	;1í	3+	3+
14	104-4	21R57	;1=	3+	33 <sup>+</sup>
15	107-1	45R35	;	3+	33 <sup>+</sup>
16	108-1	57R27	;	$33^+$	3+
17	162	93R7	1 <sup>-</sup> 1	3+	3+
18	162A	93R15	1	$33^+$	33 <sup>+</sup>
19	162-1	93R47	;1=	33 <sup>+</sup>	3+
20	162-2	93R39	;1 <sup>-</sup>	3+	33 <sup>+</sup>

with resistant pustules) against pathotype 77-5 and TR (Trace i.e. <1% severity with resistant pustules) against pathotype 104-2 during 2 years of testing. In comparison, check Agra Local produced susceptible response of 80S (80% severity of rust with susceptible pustules) to pathotype 77-5, and 70S to pathotype 104-2, while Kharchia Local produced susceptible response of 90S to pathotype 77-5 and 70-80S to pathotype 104-2 (Table 2). Results further suggest that leaf rust resistance of Synthetic 45 is also highly effective at adult stage of plant growth against the most prevalent pathotypes (77-5 and 104-2) in India. The reaction on  $F_{1s}$ , Synthetic 45 x Agra Local and Synthetic 45 x Kharchia Local produced susceptible ITs '33<sup>+</sup>', '3<sup>+</sup>', respectively (Fig. 1) indicating that resistance in Synthetic 45 is recessive in nature (Table 3).

Table 2.Adult plant response of Synthetic 45, Agra Local<br/>and Kharchia Local against leaf rust (*Puccinia*<br/>*triticina*) pathotypes 77-5 and 104-2 in field<br/>conditions during 2014-15 and 2015-16

Pathotype Year		Parental line					
		Synthetic 45	Agra local	Kharchia local			
77-5	2014-15	5R	80S	90S			
	2015-16	5R	80S	90S			
104-2	2014-15	TR	70S	80S			
	2015-16	TR	70S	70S			

The two F<sub>2</sub> populations derived from the crosses, Synthetic 45 × Agra Local (266 plants) and Synthetic 45 x Kharchia Local (135 plants) tested against pathotype 77-5 segregated into 1 resistant: 3 susceptible ratio with good fit ( $\chi^2$ =0.1253, P<sub>1d.f.</sub> =0.7234 and  $\chi^2$ =0.0617, P<sub>1d.f.</sub>=0.8038, respectively) suggesting monogenic recessive genetic control of resistance. In addition, the 47 plants of  $BC_1F_1$ population generated by Synthetic 45/Agra Local// Synthetic 45 and 42 plants of BC<sub>1</sub>F<sub>1</sub> population generated by Synthetic 45/Kharchia Local//Synthetic 45 fitted well in test cross ratio of 1R:1S ( $\chi^2$ =0.1915,  $P_{1d.f.}=0.6617$  and  $\chi^2=0.3809$ ,  $P_{1d.f.}=0.5371$ respectively) confirming the F<sub>2</sub> results of functioning of single recessive gene. BC1F1 populations derived from backcrossing of F<sub>1s</sub> with susceptible parent, Synthetic 45/Agra Local//Agra Local (157 plants) and Synthetic 45/Kharchia Local//Kharchia Local (92 plants) did not segregate into resistant and susceptible phenotypes but all seedlings tested showed susceptibility to pathotype 77-5, as expected. The genetic analysis of both the populations demonstrated and supported that the resistance in Synthetic 45 is governed by a single recessive gene. The resistance gene identified in Synthetic 45 has been putatively named as 'LrSyn45'.

# Identification of molecular marker(s) linked to the leaf rust resistance in Synthetic 45

Out of the 807 SSR markers, 115 showed

Table 3.	Infection types on parental lines, F <sub>1</sub> , F <sub>2</sub> and BC <sub>1</sub> F <sub>1</sub> of crosses involving Synthetic 45, Agra Local and Kharchia
	Local against pathotype 77-5 of <i>P. triticina</i> at seedling stage at mean temperature range 20-28 <sup>0</sup> C

Line/population	No. seedlings			Expected ratio	$\chi^2$	df	P-Value
	Resistant	Susceptible	Total	(R:S)			
	(IT ;1 <sup>=</sup> to 1 <sup>+</sup> )	(IT 3 to 3 <sup>+</sup> )					
Synthetic 45 (Syn. 45)	12	0	12				
Agra Local	0	10	10				
Kharchia Local	0	11	11				
Syn.45/AL-F <sub>1</sub>	0	14	14				
Syn.45/KL-F <sub>1</sub>	0	10	10				
Syn.45/AL-F <sub>2</sub>	69	197	266	1:3	0.1253	1	0.7234
Syn.45/KL-F <sub>2</sub>	35	100	135	1:3	0.0617	1	0.8038
Syn.45/AL//Syn.45-BC <sub>1</sub> F <sub>1</sub>	22	25	47	1:1	0.1915	1	0.6617
Syn.45/KL//Syn.45-BC <sub>1</sub> F <sub>1</sub>	19	23	42	1:1	0.3809	1	0.5371
Syn.45/AL//AL-BC <sub>1</sub> F <sub>1</sub>	0	157	157	0:1			
Syn.45/KL//KL-BC <sub>1</sub> F <sub>1</sub>	0	92	92	0:1			

polymorphism between the two parental lines, Synthetic 45 and Agra Local. The polymorphic markers were used for Bulked Segregant Analysis (BSA). Out of 266 transplanted  $F_2$  plants, only 134  $F_2$  plants survived and were considered for DNA isolation and molecular mapping of leaf rust resistance gene. BSA performed with 115 polymorphic SSR markers on contrasting bulks along with parents showed two SSR markers, *Xwmc432* and *Xcfd15* located on short arm of chromosome 1D as co-segregating with leaf resistance gene in Synthetic 45 and were recognized as putatively linked (Fig. 2). The  $F_2$  population was



# Fig. 2. Bulked Segregant Analysis by SSR markers *Xwmc432* and *Xcfd15* located on short arm of chromosome 1D

genotyped using both the co-segregating SSR markers, *Xwmc432* and *Xcfd15*. The calculated value of Chisquare tests for SSR marker, *Xwmc432* ( $\chi^2_{(1:2:1)}$  =2.999, P<sub>2df</sub>=0.2231) and *Xcfd15* ( $\chi^2_{(1:2:1)}$  =0.7313, P<sub>2df</sub>=0.6937) individually fit well in the expected genetic ratios in F<sub>2</sub> populations. The F<sub>2</sub> phenotypic classes (resistant and susceptible) were used for genetic linkage analysis. Linkage was detected between the *LrSyn45* and the SSR marker *Xwmc432* ( $\chi^2_L$ =136.6817, P<sub>2df</sub><0.001) (Table 4). Similarly, the linkage was detected between *LrSyn45* and SSR marker *Xcfd15* ( $\chi^2_L$ =101.8559, P<sub>2df</sub><0.001) (Table 5).

Table 4.Joint segregation of co-dominant SSR markerXwmc432 with F2 phenotypes from the crossSynthetic 45 x Agra Local

F <sub>2</sub> Phenotype	Xwmc432			32	$\chi^{2}_{(1:3)}$	P-Value	
	A	Н	В	Total			
Resistant (rr)	37	1	3	41	2.2388	1df,0.1346	
Susceptible (R-)	2	56	35	93			
Total	39	57	38	134			
$\chi^2$ (1:2:1) = 2.9999, 2df, P = 0.2231							
$\chi^2 (\mbox{Compounded})$ 3:6:3:1:2:1=141.9204, 5df, P<0.001							
$\chi^2$ difference (Linkage) = 136.6817, 2df, P<0.001							

A=Homozygous for Synthetic 45 allele, B=Homozygous for Agra Local allele, H= Heterozygous

Table 5.	Joint segregation of co-dominant SSR marker						
	Xwmc432 with $F_2$ phenotypes from the cross						
	Synthetic 45 x Agra Local						

F <sub>2</sub> Phenotype		Хс	fd15	5	χ2 <sub>(1:3)</sub>	P-Value	
	A	Н	В	Total			
Resistant (rr)	33	7	1	41	2.2388	1df,0.1346	
Susceptible (R-)	4	60	29	93			
Total	37	67	30	134			
$\chi^2$ (1:2:1) = 0.7313, 2df, P = 0.6937							
$\chi^{2}$ (Compounded) 3:6:3:1:2:1= 104.8258, 5df, P<0.001							
$\chi^2$ difference (Linkage) = 101.8559, 2df, P<0.001							

A=Homozygous for Synthetic 45 allele, B=Homozygous for Agra Local allele, H= Heterozygous

Linkage ( $\chi^2_L$ =189.4333, P<sub>4df</sub><0.001) was also detected between the two SSR markers, *Xwmc432* and *Xcfd15*.

The  $\chi^2$  linkage value between *LrSyn45* and the marker Xwmc432 is higher than the  $\chi^2$  linkage value between LrSyn45 and the marker Xcfd15 suggesting that LrSyn45 is closer to the marker Xwmc432 as compared to Xcfd15. The highest  $\chi^2$  linkage value between the two SSR markers showed that two markers are at the closest distance in comparison to the distances between LrSyn45 and the two markers. This analysis indicated the sequence of gene and the markers as LrSyn45-Xwmc432-Xcfd15. The result of linkage analysis using software MAPMAKER version 3.0 (Lander et al. 1987) for construction of linkage map suggested that the leaf rust resistance gene in LrSyn.45 is located at 6.1 cM distal to the marker Xwmc432 and 10.6 cM distal to another marker Xcfd15 (Fig. 3). The genetic distance between the two markers



Fig. 3. Linkage map of leaf rust resistance gene LrSyn45

was 4.6 cM. Linkage map also suggested the sequence of gene and the markers as *LrSyn45-Xwmc432-Xcfd15*.

Among the known leaf rust resistance gene, three genes viz., Lr21, Lr42 and Lr60 have been reported to be located on short arm of chromosome 1D. LrSyn45 was considered different from Lr21, because Lr21 is ineffective to Indian pathotypes 12, 12B, 77, 77A-1, 104B and 162 of leaf rust pathogen (Kumar et al. 1988), whereas in our study, Synthetic 45 showed resistance to a common pathotype 162 and other pathotypes of race groups 12, 77 and 104. Regarding the possibility of LrSyn45 being the Lr42 is not clear because of different reports on the nature of inheritance of Lr42 viz., dominant (Czembor et al. 2008), partial dominant (Cox et al. 1994) and recessive (Liu et al. 2013). However, the inheritance of proposed resistance gene, LrSyn45 is recessive in the present study. Another leaf rust resistance gene Lr60 (LrW2), also reported to be located on chromosome1DS is dominant in nature (Hiebert et al. 2008). The uniqueness of LrSyn45 in synthetic 45 suggests the identified resistance is presumably diverse. However, for further confirmation of the uniqueness of the gene, the test of allelism with *Lr42* and *Lr60* and fine mapping of *LrSyn45* is required.

# Authors' contribution

Conceptualization of research (JBS, PCG, V, SKJ); Designing of the experiments (PCG, JBS, V, SKJ); Contribution of experimental materials (JBS); Execution of field/lab experiments and data collection (PCG); Analysis of data and interpretation (PCG, JBS, V, NM, SKJ); Preparation of manuscript (PCG, JBS, V, NM, SKJ)

# Declaration

The authors declare no conflict of interest.

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