Inheritance studies for morphological characters and sex expression in pistillate lines of castor (Ricinus communis L.)

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Abstract

Inheritance of morphological characters in four crosses of pistillate x pistillate lines indicated that stem colour was controlled by single gene while red stem colour was dominant over green stem colour. However, green stem colour in the cross DPC 13 x M 574 and green colour of petiole, capsules and spines in cross M 574 x DPC 11 showed digenic epistatic ratio to red stem colour, red colour of petiole, capsules and spines. The characters plant type and bloom were monogenically controlled while normal plant type and presence of bloom were dominant over dwarf plant type and absence of bloom respectively. Joint segregation analysis indicated that stem colour, plant type and bloom were segregating independently in all the four crosses. Pistillate character was dominant over pistillate with interspersed staminate flowers [ISF] and $\mathsf{controlled}$ by one $[\mathsf{P}_1]$ or two $[\mathsf{P}_1\mathsf{P}_2]$ to four genes $[\mathsf{P}_1\mathsf{P}_2]$ **P3P4]. Joint segregation analysis indicated linkage between stem colour and dwarf plant type in the cross DPC 13 x DPC 11 and pistillate character [P₁] and plant type [PtN] in cross M 574 x DPC 11. The distance between stem colour [Rst/gst] to plant type [PtDw] and single gene for pistillate character and plant type was estimated as 40.2 centimorgans and 47.6 centimorgans based on square root** of frequency of double recessive phenotypes in **F₂ data.**

Key words: Castor [Ricinus communis L.,], sex expression, morphological characters, independent segregation and linkage

Introduction

Morphological characters like colour of stem, petiole, capsule, spines, rachis, midrib, bloom, plant type, spike type, number of nodes to primary spike are important markers used for distinctness, uniformity and stability (DUS) tests in castor (Ricinus communis L.) under different environments. Stem colour in castor is classified as red, green and mahogany based on the pigmentation of sap colour in epidermal palisade cells and parenchymatous areas of the stem [1]. Majority of the pistillate lines which are derivatives of the VP-1 back

ground have dwarf plant type with condensed internodes and cup shaped leaf and serve as distinct morphological markers and facilitate rouging in seed production plots. However, information on inheritance of morphological markers viz., stem colour, plant type, and arrangement of nodes on the stem, leaf shape etc., and their linkage has not been worked out and the available information is either scanty or very old. Castor, a sexually polymorphic species produces pistillate spikes with interspersed staminate flowers (ISF), monoecious or revertants [2]. Pistillate lines are maintained by interspersed staminate flowers (ISF) called refined or modified method of seed production of pistillate lines. The production of these interspersed staminate flowers is highly environmentally sensitive [3]. Information on inheritance of genes for ISF and interaction with other morphological characters in pistillate lines is less worked out. The present study is an attempt to study the inheritance of morphological characters and sex expression in four crosses of pistillate x pistillate lines of castor.

Materials and methods

Three pistillate lines viz., DPC 9, DPC 11, DPC 13 developed by conventional breeding methods and two pistillate lines viz., M 571, M 574 developed by gamma ray (55 Kr) treatment of VP-1 seeds [4] differing in morphological characters like stem colour, bloom, plant type, spike nature, duration to maturity were chosen for the present study [Table 1]. Four crosses viz., DPC 9 x M571, DPC 13 x DPC 11, DPC 13 x M 574, M 574 x DPC 11 were made in Rabi 1999. The F₁s were raised in Rabi 2000, selfed and back crossed to both of their parents. All the three generations viz., P_1 , P_2 , F_3 , F_4 , F_5 , B_1 and B_2 were planted in R abi 2001 and observations on both morphological characters and sex expression were recorded on individual plants in each generation. Observations on stem colour were recorded in all

 populations of four crosses while colour on other external parts like petiole, capsules, spines, rachis, midrib were recorded in progeny of the cross IV-M 574 x DPC 11 only. Observations on \geq 20 plants were recorded on parents, F_1 s, B₁, B₂ while sample size in F_2 varied in different crosses. The chi-square test was applied for the goodness of fit for different expected genetic ratios. Joint segregation analysis and linkage analysis was done based on the formulae for the estimation of linkage from $F₂$ data [4].

Results and discussion

The inheritance of stem colour was studied in all the four crosses (Table 2). In three crosses involving either green x red (DPC 9 x M 571) or red x green (DPC 13 x DPC 11) or green x green (M 574 x DPC 11) red stem colour is dominant over green in F_1 . The F_2 segregation ratios in three crosses indicated that stem colour is monogenic and segregated in to 3:1 ratio in all the three crosses mentioned above. A single gene Rst/rst is responsible for stem colour in three crosses of castor. Monogenic inheritance for stem colour and dominance of coloured stem over green stem was reported in earlier studies [5-7]. However, in cross DPC 13 x M 574, green stem colour is dominant over red stem colour in $\mathsf{F}_{_{1}}$ and F_2 population indicated a good fit to the digenic epistatic

ratio of 9 green: 6 red: 1 mahogany where green stem colour is epistatic to red stem colour. The presence of either of two genes Rst or Gst individually produces red stem colour while presence of both the two genes RstGst polymerizes the effect to turn to green colour. The presence of two recessive genes *rstgst* produces mahogany stem colour in cross DPC 13 x M 574. Mahogany colour has both green and red colours with predominance of red colour. Epistatic ratios for stem colour were also reported [7, 8]. Epistatic interaction of two factors M and G resulted in three colours where factor G converts tinged green to green and mahogany to rose and M is the factor for mahogany [6].

The inheritance of colour on petiole, capsules, rachis, spines and midrib was studied in the cross M 574 x DPC 11 (Table 2). Petiole colour is governed by digenic supplementary epistatic gene action and $F₂$ population fits in 9 red: 4 green:3 mahogany ratio. A gene gpt has no effect of its own but modifies the action of Rpt which produces mahogany petiole colour. Presence of both the dominant genes Rpt-Gpt- produces red petiole colour while rptrpt Gpt- produces green petiole colour and Rpt-gptgpt mahogany stem colour.

Capsule colour is predominantly green in the cross M 574 x DPC 11 and $F₂$ population segregated to 15

Parents /crosses	Stem color	Bloom	Plant type	Leaf shape	Spike type*	Duration	No. of nodes to primary
Cross-1							
DPC ₉	Green	Zero	Normal	Flat	SC	Early	9
M 571	Red	Triple	Dwarf	Cup	L	Medium	10
DPC 9 x M 571 (F ₁)	Red	Double	Normal	Flat	L	Early	$\,8\,$
Cross _{II}							
DPC 13	Red	Zero	Dwarf	Cup	C	Medium	11
DPC 11	Green	Double	Normal	Flat	L	Medium	14
DPC 13 x DPC 11 (F_1)	Red	Double	Normal	Flat	SC	Early	$\boldsymbol{9}$
Cross III							
DPC 13	Red	Zero	Dwarf	Cup	C	Medium	15
M 574	Green	Triple	Dwarf	Cup	SC	Late	17
DPC 13 x M 574 (F_1)	Red	Triple	Dwarf	Cup	SC	Late	16
Cross IV							
M 574	Red	Triple	Dwarf	Cup	SC	Late	17
DPC 11	Green	Double	Normal	Flat	L	Medium	14
M 574 x DPC 11 (F ₁)	Red	Triple	Normal	Flat	SC	Medium	13

Table 1. Pigmentation and morphological character states in 5 parents and the F₁s in 5 crosses of castor

 ${}^{\ast}C$ = compact, SC = Semi compact, L = Loose

green:1 red capsule colour indicating the role of either of the duplicate genes Rcp or Gcp and both the genes producing green capsules while recessive duplicate genes rcp/gcp result in red colour. Similar ratios have been reported earlier [1]. However, monogenic inheritance of capsule colour and dominance of green colour was reported in other studies [9, 10].

In the same cross M 574 x DPC 11, colour of spines on the capsule was recorded as red, green and mahogany and $F₂$ population fit to the epistatic gene ratio of 12 red : 3 green: 1 mahogany. Two genes Rsp and Gsp control two colours- red and green spine colours while the presence of dominant Rsp masks the effect of Gsp and gsp resulting in Rsp- Gsp- as red colour while rsp rsp Gsp- produces green spine colour and absence of both the dominant genes (rspgsp) resulted in mahogany spine colour.

Unlike the other parts, midrib of castor leaves is of two colours red and white and the F_2 population indicated the role of trigenes and fit into the ratio of 15 red: 49 white while red is dominant over white midrib colour.

Bloom or waxy coating in castor is an important morphological marker and serves as a natural protection against drought, cold, jassids etc., Castor plants are mainly classified as bloom or no bloom based on the presence or absence of bloom on external parts of the plant. However, based on the presence of bloom on combination of plant parts, genotypes in castor were usually classified as single bloom (stem + petiole + capsule stalks), double bloom [stem, petiole, capsule + lower side of the leaf] and triple bloom (all the above parts + upper side of the leaf) [1]. Though the data on different bloom types were recorded in segregating generations of all the four crosses, none of the data could fit into any genetic ratios. Differences in bloom distribution on different plant parts may be due to variation in the penetrance and expressivity of the genes controlling the bloom character or due to multiple alleles. In the present study, inheritance of bloom character indicated that presence of bloom is dominant over no bloom in F_1 and F_2 population has a good fit to 3:1 ratio (Table 3). In cross, M 574 x DPC 11, as both the parents M 574 [triple bloom] and DPC 11 (double bloom) are bloom types, F₂ population is classified as triple vs either single or double and fit to the 3: 1 ratio indicating the role of single gene [PtBl] for bloom character. Similar monogenic inheritance for bloom character was reported by earlier authors [6, 7, 8, 11]. Epistatic interaction was reported in a cross between double bloom and no bloom

Table 2. Inheritance of stem color in four crosses of castor and color of petiole, rachis, spine and midrib in cross 4

ration					Cross/gene- No. of plants observed Expected χ^2 ratio value
	Total		Red Green Maho-	gany	
			Stem color		
Cross-I (DPC 9 x M 571)					
F_{2}	66	52	16	0	3:1 0.15
$F1$ x DPC 9 22		12	10		1:1
F_1 X M 571 20		20			
Cross II (DPC 13 x DPC 11)					
F_{2}	190	142	48	0	3:1 0.007
F_1 x DPC 13 26		14	12		1:1
F ₁ x DPC 11 24			24		
Cross III (DPC 13 x M 574)					
F_{2}	170	61	99	10	6:9:1 0.209
F ₁ x DPC 13 22		7	8	6	1:1:1
F ₁ x DPC 11 21				21	
Cross IV (M 574 x DPC 11)					
F_{2}	197	146	51		3:1 0.11
$F_1 x M 574 20$		20			
F ₁ x DPC 11 20		11	10		1:1
	Petiole color				
$F_{\rm 1}$	20	20			
F ₂	92	99	55	38	$9:4:3$ 1.88
F_1 x M 574	20	20			
F ₁ x DPC 11 20		7	7	6	1:1:1
		Capsule color			
F_{1}	20	20			
F ₂	192	16	176		15:1 1.419
$F_1 x M 574 20$		20			
F_1 x DPC 11 20 11 10		Spine color			1:1
F_{1}	21		21		
F ₂	153	10 [°]	114	29	12:3:1 0.119
$F_1 x M 574 20$		$\overline{7}$	8	- 5	
F, x DPC 11 20			20		
		Midrib color			
			Red White		
$F_{\rm i}$	20	20			
F ₂	124	32	92		15:49 0.405
$F_1 \times M 574 20$		20			
F_1 x DPC 11 20		11	10		1:1

and F_2 ratio of 9 double boom: 3 single bloom: 4 no bloom was due to epistatic effect of factor B over factor C and factor C has no role on its own and it merely intensifies the action of factor B [12].

Majority of the pistillate lines derived from VP-1 back ground have dwarf stem, condensed nodes, cup shaped leaves and convergent branching which are highly linked characters [13]. In two crosses, DPC 9 (N) x M 571 (Dw) and M 574 (Dw) x DPC 11(N) involving normal (N) and dwarf (DW) plant types, normal plant type is dominant over dwarf in F_1 and F_2 segregated in to 3 normal : 1 dwarf indicating the role of single gene – PtN as reported earlier [14, 15 and 11] (Table 3). The $F₁$ population of the cross -DPC 13 (Dw) x DPC 11 (N), indicated that normal plant type is dominant over dwarf plant type. The $F₂$ segregation fits in to 45:19 ratio indicating the role of three genes for plant type. However, in the cross III involving both dwarf type parents – DPC 13 and M 574, dwarf plant type is dominant over normal plant type and $F₂$ population indicated the role of duplicate dominant genes – either PtDw or PtN producing the same plant type – dwarf while duplicate recessive genotype – ptdw / ptn results in normal plant type.

In castor, spikes are classified based on the arrangement or density of capsules on the spike as Loose, Compact and Semi compact [1]. Compact and

Semi compact spikes are highly susceptible to fungal diseases like Botrytis under conditions of high humidity, rainfall, cloudy weather due poor aeration and ventilation. Earlier studies on inheritance pattern [5] indicated incomplete dominance and $F₂$ populations had all the patterns of spike nature. In the present study also, the distinction between semi compact and compact types is not clear as majority of the pistillate lines had medium long or long compact/semi compact spikes except the genotype M 571, which has a loose spike. In a cross between compact (DPC 13) x semi compact (M 574), single gene SpSc controlled the compactness of spike and F_2 population segregated in to 3 semi compact : 1 loose spike indicating the dominance of semi compact spike over loose spike. In cross II, DPC 13 x DPC 11, two epistatic inhibitory genes controlled the spike type while F_2 population fit into the ratio of 13 semi compact: 3 compact spike types. The presence of dominant gene at one locus (SpSc) and recessive gene at other locus (Spll) produced the same phenotype – semi compact spike while presence of dominant gene at other locus (spL) resulted in loose spike types. The $F₂$ population in cross IV indicated test cross ratio of 1 loose: 1 semi compact spike types (Table 4).

Number of nodes to primary spike in castor indicates the duration of the flower initiation of the primary spike and on an average every node takes 4

Table 3. Inheritance of bloom and plant type in four crosses of castor

Cross/generation	No. of plants observed		Expected ratio	χ^2 value		No. of plants observed Expected				χ^2 value	
	Total	Loose	SC/ compact			Total	Early	Medium Late			
Cross II							(<12N)	$(13-16N) > 17N$			
F ₂	154	26	128	3:13	0.382	143	75	60	8	9:6:1	1.35
$F1$ x DPC 13	26		26			26	10	8	8	1:1:1	
$F1$ x DPC 11	24	12	12	1:1		24	24				
Cross IV											
F ₂	196	89	107	1:1	1.652	196	128	68		45:19	2.45
$F1$ x DPC 11	20	9	11	1:1		20	11	9			
$F_1 \times M$ 574	20		20			20	20				
Cross III											
F ₂	162	43	119	1:3	0.258						
$F1$ x DPC 13	22		22	1:1							
$F_1 \times M$ 574	21	11	10								

Table 4. Inheritance of Spike type in cross II, III, IV and node number in cross II and IV

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days to develop [14]. However, node number varies within and among genotypes based on the season of planting and location and within a location; node number of a genotype is constant for each planting season and hereditary [15]. Based on the previous records at Directorate of Oilseeds Research, Hyderabad, genotypes were classified as early (<12 nodes), medium (13-16 nodes) and late (>17 nodes) based on the node number. Inheritance of node number indicated that in a cross between medium x medium (DPC 13 x DPC 11), F_1 was early and F_2 population segregated in to 9 early: 6 medium: 1 late ratio. The presence of two dominant genes $(N, -N, -)$ results in low node number (<12) while the presence of dominant gene at the second locus (n₁- N_{2} -) results in medium node number (13-16N) while the double recessive genotype n₁n₁n₂n₂ produces higher node number (>17N). In cross III, inheritance of node number indicated that three genes- $n_1n_2n_3$ control the character and F₂ population segregates to inhibitory epistatic ratio of 45 early : 19 medium types (Table 4).

Sex expression in castor is polymorphic and varies from monoecious, pistillate, ISF and revertants. A pistillate spike with staminate flowers interspersed in between the female flowers from the base to the top is termed as Interspersed staminate flower (ISF). In the present study, pistillate lines are crossed with interspersed staminate flowers of other pistillate lines. The F_1 s in all the four crosses were pistillate with production of occasional ISF in higher temperatures

indicating the dominance of pistillate over pistillate lines with ISF. Sex expression in $F₂$ population was recorded in both primary and secondary spikes in all the four crosses (Table 5). The number of genes controlling pistillate expression varied from one (P_1) to two genes (P_1P_2) in cross I and cross IV to three genes $(P_1P_2P_3)$ and four genes $(P_1P_2P_3P_4)$ in cross II and III. In all the cases pistillate expression is dominant over pistillate with ISF as reported by earlier authors $[13, 3]$. The F₂ population segregated to simple monogenic ratios like 3 pistillate:1 pistillate with ISF (in cross I) or 1 pistillate: 1 ISF (cross IV) or digenic epistatic ratios like 13 pistillate:3 ISF in secondary of cross IV. In cross II, $F₂$ population was a good fit to tetragenic epistatic ratios in primary (162:94) and secondary (229: 27). The change in the segregation pattern of secondary spikes denotes the interaction of genotype with environment viz., daily maximum temperatures, day length etc. The interaction of sex expression with environmental conditions is well documented earlier [2, 3]. More number of genes was activated along with the production of spikes to cope up with the changing environmental conditions. The role of modifying genes was indicated in the earlier study of the author [3]. However, earlier report [3], indicated that gene $F₁$ for monoecious controls a genetically stable series of sex variants ranging from female 'f' to strong male inbreds and sex expression of such strains is controlled by non genetic variations which occur intermittently during the development of individual plants.

		Generation Obs/exp No. of plants Primary			No. of plants Secondary	χ^2 value	
		ISF Pistillate			Pistillate ISF		
Cross I	DPC 9 x M 571						
F_{2}	(O) (E) Ratio	52 49 3 $\ddot{\cdot}$	14 17 1				0.51
B_1	(O) (E)	12 11	10 11				
Genetic ratio used		$\ddot{\cdot}$ 1	1				
B ₂	(O) (E)	20 20					
Cross II	DPC 13 x DPC 11						
	(O) (E) Ratio	102 104 $\ddot{\cdot}$ 162	62 60 94	:	131 131 229	16 16 27	0.105 0.0
B_1	(O) (E)	26 26					
B ₂	(O) (E)	24 13	11		24 14	10	
Genetic ratio used		1 $\ddot{\cdot}$	1	$\ddot{\cdot}$	1	1	
Cross III	DPC 13 x M 574						
	(O) (E) Ratio	157 158 $\ddot{\cdot}$ 63	4 3 1		112	0	0.339
B ₁	(O) (E)	22 22					
B ₂	(O) (E)	14 12	10 12				
Genetic ratio used		1	1				
Cross IV	M 574 x DPC 11						
	(O) (E) Ratio	90 94 $\ddot{\cdot}$ 1.	99 95 1		154 155 13	37 36 $\ddot{\cdot}$ 3	0.338 0.033
B_1	(O) (E)	20 20					
B ₂	(O) (E)	12 10	8 10				
Genetic ratio used		1 :	1				

 Table 5. Pattern of segregation for Pistillate vs ISF in segregating material of four crosses of castor

Table 6. Pattern of joint segregation for stem color vs bloom in crosses I, II and IV

Generation Genetic	ratio used		No. of plants					
					Stem colour vs. bloom		value	
Cross I	DPC 9 x M 571 RBI			GBI	RNbl	GNbl		
$\overline{\mathsf{F}_2}$	9:3:3:1	(O)	37	14	11	4	0.38	
		(E)	37	12 [°]	12	4		
$\mathsf{B}_{\scriptscriptstyle{1}}$	1:1:1:1	(O)	5	4	6	5		
		(E)	5	5	5	5		
B ₂		(O) (E)	20 20					
Cross II	DPC 13 x DPC 11							
F_{2}	9:3:3:1	(O)	108	34	41	7		
		(E)	106	36	36		12 2.95	
B_{1}	1:1	(O)	26	24				
		(E)	25	25				
B ₂	1:1	(O)	12		8			
		(E)	10		10			
Cross IV	M 574 x DPC 11							
F_{2}	9:3:3:1	(O)	110	36	34	11		
		(E)	107	36	36		12 0.274	
B_{1}		(O)	22					
		(E)	22					
B ₂	1:1:1:1	(O)	8	9	7	7		
		(E)	8	8	8	8		

 $RBI = Red$ bloom, $GBL = Green$ no bloom, $RNbl = Red$ no bloom, GNbl = green no bloom

Table 7. Pattern of joint segregation for stem color vs plant type in crosses I, II and IV

Gene- ration	Genetic ratio used			No. of plants				
		O/E	RN	Rdw		GN Gdw		
Cross I	DPC 9 x M 571							
$\mathsf{F}_{_2}$	9:3:3:1	(O) (E)	44 37	7 12	11 12	4 4	3.48	
B_{1}		(O) (E)	11 10			9 10		
B_2		(O) (E)	12 10	8 10				
Cross II	DPC 13 x DPC		11					
F_{2}	135:57: (O) 45:19	(E)	92 100	50 34	31 42	14	17 11.69**	
B ₁	1:1	(O) (E)	11 10	9 10				
B_{2}	1:1	(O) (E)	11 12		13 12			
Cross IV	M 574 x DPC 11							
$\mathsf{F}_{_2}$	9:3:3:1	(O) (E)	105 110	36 37	38 37	17 12	2.37	
B ₁	1:1	(O) (E)	12 ¹² 10	8 10	11 10	9 10		
B_2		(O) (E)			20 20			

 $RN = Red$ normal, $Rdw = Red$ dwarf, $GN = Green$ normal, Gdw = green dwarf

Genetic ratio used 1 : 1 Joint segregation analysis for stem colour and :

bloom and stem colour vs plant type in three crosses – I, II, IV (Table 6 and Table 7) indicated independent segregation of stem colour and bloom which are controlled by single genes $-$ Rst /gst and BI/bl. In case of stem colour vs plant type also, independent segregation of characters was observed in cross I, II and IV where the characters were controlled by single genes Rst/gst and Dw/dw and no linkages were obtained. However, in cross II, DPC 13 x DPC 11, stem colour red is linked to dwarf plant type based on the deviation of χ^2 from the estimated 9:3:3:1 ratio. Linkage is estimated based on the square root method of

Table 8. Joint segregation in cross III for stem color Vs plant type

Gener- ation	Observed/ expected	No. of plants						χ^2 value
				GDw RDw MDw GN		RN	MN	
F_{2}	(O) (E) Ratio	99 90 135	56 60 90	8 10 15	1 5 9	5 4 6	2 1 1	7.02
$B_{\scriptscriptstyle 4}$	(O) (E) Ratio	12 12 1	11 12 1	13 12 1				
B_{2}	(O) (E)	25 25						

frequency of double recessive non cross over gametes in the case of coupling phase type of linkage [16]. The distance between the locus for stem colour (Rst/gst) to that for plant type (PtDw) is estimated as 40.2 centimorgans. Joint segregation of stem colour and plant

Table 9. Joint segregation in cross III for stem color Vs bloom

ation	Gener- Observed/ expected			value				
		GBI			RBI MBI GNbIRNbI MNbI			
F_{2}	(O) (E) Ratio	73 72 27	44 48 18	15 8 3	25 24 9	12 16 6	1 2 1	2.76
BC,	(O) (E)	7 5	6 5	4 5	5 5	5 5	3 5	
Ratio		1	1	1	1	1	1	
BC,	(O) (E) Ratio	11 10 1	9 10 1	10 10 1				

 $G =$ Green, $R = Red$, $M =$ Mahogany, $Dw = Dw$ arf, $N =$ Normal $G =$ Green, $R =$ Red, $M =$ Mahogany; BI = Bloom, NBI = No bloom

Table 10. Joint segregation for plant type and sex expression in segregating material of four crosses of castor

 type and stem colour and bloom (Tables 8 and 9) in cross III with its six phenotypic classes unlike the other three crosses also indicated independent segregation of the two genes controlling each of the character-stem $\mathsf{colour}\;(\mathsf{Rst}_{\mathsf{1}}\mathsf{Rst}_{\mathsf{2}}/\mathsf{gst}_{\mathsf{1}}\mathsf{gst}_{\mathsf{2}})$ and bloom $(\mathsf{Bl}_{\mathsf{1}}\mathsf{Bl}_{\mathsf{2}}/\mathsf{bl}_{\mathsf{1}}\mathsf{bl}_{\mathsf{2}}).$

Pistillate character has been introduced along with TSP 10 R, an exotic pistillate line with dwarf, condensed internodes and cup shaped leaves [3]. Thus joint segregation analysis for plant type and pistillate expression was studied in both primary and secondary spikes of three crosses (Table 10). In two crosses, II and III, plant type and pistillate expression had independent segregation between double (Dw_1Dw_2) / triple genes ($Dw_{_1}Dw_{_2}Dw_{_3}$) of plant type and three (P₁P₂ P_3) and four ($P_1P_2P_3P_4$) genes for pistillate expression. However, in cross IV, monogenic genes for plant type and sex expression were linked as indicated by the frequency of double recessive non cross over gametes as the characters were linked in coupling phase. Linkage between genes for plant type and sex expression was estimated as 47.6 centimorgans based on square root of frequency of double recessive phenotypes of F_2 data [16]. Presence of linkage between morphological characters like stem colour and plant type and plant type and sex expression has not yet been reported.

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References

- **1. Kulkarni L. G. and Ramanamurthy G. V.** 1977. Castor, ICAR, New Delhi.
- **2. Shiffriss O.** 1960. Conventional and unconventional systems controlling sex variations in Ricinus. J. Genetics, **57**: 573-578.
- **3. Lavanya C.** 2002. Sensitivity and variations of sex expression in response to environmental changes in castor. Ind. J. Gen. Pl. Breed., **62**: 232-237.
- **4. Richharia R. H., Ghosh A. K., Prakasa Rao S. V. S. and Misro B.** 1966. Bulletin No. 5. Formulae for the estimation of linkage from $\mathsf{F}_2^{}$ data. CRRI, Cuttack, India.
- **5. White O. E.** 1918. Inheritance studies on castor beans. Mem. Brooklyn bot. Gdn., **1**: 513-521.
- **6. Harland S. C.** 1922. Inheritance in Riciuns communis L. part II. J. Genet., **10**: 207-18.
- **7. Harland S. C.** 1928. The genetics of Riciuns communis L. Biblie. Genetica, **4**: 171-77.
- **8. Seshadri C. R. and Varisai Mohammad.** 1951. Study of inheritance of certain characters in castor. Proc. of the first scientific workers conference, Agrl. College and Res. Inst, Coimbatore.
- **9. Patwardhan G.** 1931. A preliminary note on inheritance of castor. J. Indian Bot. Soc., **10**: 100-109.
- **10. Solanki S. S. and Joshi P.** 2001. Inheritance study of morphological traits in castor (Riciuns communis L.). Indian J. Genet., **61**: 136-139.
- **11. Peat J. E.** 1926. Genetic studies in Riciuns communis L., J. Genet., **19**: 373-89.
- **12. Hanumantha Rao C., Raoof M. A. and Lavanya C.** 2005. Study on segregation patterns and linkages between morphological characters and wilt resistance in castor (Ricinus communis L.). J. Oilseeds Res., **22**: 114-118.
- **13. Zimmerman L. H.** 1958. Castor bean, a new oil crop for mechanized production. Adv. in Agron., **10**: 257- 288.
- **14. Moshkin V. A.** 1986. Castor. Oxonian Press Pvt. Ltd., New Delhi.
- **15. Masur N. G.** 1953. Castor breeding in Bombay Presidency. Agrl. Live stlk. India, **3**: 125-43.
- **16. William D. Stansfield.** 1969. Schaum's outline of theory and problems of genetics.