



Genetic basis of rust resistance of a high yielding wheat line from northern hills zone

D. Datta¹, S. C. Bhardwaj² and M. Prashar²

¹Indian Institute of Vegetable Research, Varanasi 221 305

²Directorate of Wheat Research, Regional Station, Flowerdale, Shimla 171 002

(Received: September 2006; Revised: February 2008; Accepted: February 2008)

Abstract

Genetic basis of rust resistance was investigated in a bread wheat line HS424 which is bred for Northern Hills Zone of India. The study showed that HS424 is resistant to leaf rust and stem rust pathotypes prevalent in India. At seedling stage it was resistant to stripe rust pathotypes except 46S119. However, at adult plant stage, HS424 showed only moderate susceptibility to pathotype 46S119. Low terminal disease against pt. 46S119 must be due to additional adult plant resistance factors in HS424. At least two resistance genes were involved in the resistance against leaf rust pathotypes where in resistance to pathotypes 77-5 and 77-2 was governed by single dominant gene and two independent dominant genes, respectively. Stem rust resistance was governed by three genes. Resistance to pt. 40A was due to two independent dominant genes whereas single dominant gene imparted resistance to pt. 40-1. Adult plant resistance gene *Sr2* was also postulated in HS424. Based on infection type data, inheritance study, morphological -marker, genetic linkage and molecular marker analyses it is concluded that HS424 carries genes *L24+Lr26+Sr2+Sr24+Sr31+Yr9*.

Key words: Genetic basis, rust resistance, wheat, HS424

Introduction

Apart from the inherent yield potential both biotic and abiotic stresses determine the realized yield of cultivars. Among the biotic stress factors, rusts are the most important diseases of wheat in India. Though stem rust (*Puccinia graminis f.sp. tritici* Eriks. & Henn.) samples are being detected regularly from places at high altitudes as Dalang Maidan of Lahaul Valley of Himachal Pradesh, leaf rust (*Puccinia triticina* Erikss.) and stripe rust (*Puccinia striiformis* Westend. f. sp. *tritici*) are more important in the northern hills. HS424 (CPAN3004//HPW(d1)30/HS286) is an advanced bread wheat line of All India Coordinated Wheat and Barley Improvement Project. Along with other advance lines and checks it was tested with multi-pathotypes of leaf, stem and stripe rusts at seedling and adult plant stages for two consecutive years under temperature controlled glass house and polythene house at the Directorate of Wheat

Research, Regional Station, Flowerdale. HS424 showed high resistance to leaf and stem rust pathotypes at seedling as well as adult plant stage. Genetic basis of rust resistance of HS424 is reported here.

Materials and methods

Plant material

The experimental material consisted of F_2 and F_3 generations derived from the cross Agra Local/HS424. Segregating generations were tested at seedling stage against predominant leaf rust pathotypes 77-2 and 121R63-1 and stem rust pathotypes 40A and 40-1.

Resistance tests

The F_2 and F_3 seedlings were raised in aluminum bread pans (trays) comprising ten rows with a susceptible check, Agra Local, at 7th row. Fully expanded primary leaves (7-8 days old) were inoculated with uredospores suspended in light, non-phytotoxic isoparaffinic oil (soltrol 170, Philips Chevron). Inoculations of leaf and stem rusts were carried out in temperature controlled glass house maintained at 22°C and 24°C, respectively. The inoculated seedlings were kept in a humid glass chamber for 48 hours and then transferred to benches. Infection types (ITs) were recorded 14 days after inoculation and classified according to standard methods [1] as follows: IT 0; (naught fleck); (fleck) and ;2 (fleck two), were classified as the resistant reactions whereas ITs 3 (three) and 3+ (three plus) were designated as susceptible reactions. The F_2 seedlings from the single F_1 plant were scored for infection types (ITs) and transplanted in the field for obtaining of F_3 families. About twenty five seedlings were analyzed per family. Same set of F_3 families were evaluated wherever progeny testing was done simultaneously with more than one pathotype. Chi-square test was applied to test the validity of expected genetic ratios. The stem rust resistance gene *Sr2* was postulated on the basis of microchlorosis [2], a phenotypic marker that is completely linked with the resistance gene *Sr2* which

Table 2. Segregation of seedling reaction against leaf, stem and stripe rust pathotypes in F₂ and F₃ generations from cross of HS424 with Agra Local

Pathotypes and generation	Number of seedlings/families			Expected ratio	χ^2 value	P range
	Resistant	Segregating	Susceptible			
Leaf rust						
Pathotype 77-2						
F ₂	335		27	15R:1 S	0.73	0.25-0.50
F ₃	33	44	6	7R:8seg:1S	0.58	0.50-0.75
Pathotype 77-5						
F ₂	301		93	3R:1 S	0.45	0.50
F ₃	19	46	28	1R:2seg:1S	1.75	0.25-0.50
Stem rust						
Pathotype 40A						
F ₂	284	76	18	15R:1 S	0.09	0.75-0.90
F ₃	41	54	5	7R:8seg:1S	0.74	0.50-0.75
Pathotype 40-1						
F ₂	293		86	3R:1S	1.14	0.25-0.50
F ₃	29	52	17	1R:2Seg:1S	3.30	0.10-0.25

resistant and 18 were susceptible (Table 2). The data gave good fit to 15 resistant: 1 susceptible ratio ($P = 0.75-0.90$). Among the 100 F₃ families, 54 segregated, 5 were homozygous susceptible and 41 were homozygous resistant (Table 2). The F₃ family segregation was good fit to 7 resistant: 8 segregating : 1 susceptible ($P = 0.50-0.75$).

Seedling resistance test with 40-1 of stem rust

The F₁'s of the cross between Agra Local and HS424 were resistant. The F₂ segregated for two infection types (2 to 2⁻ and 3⁺). Out of 379 seedlings, 293 were resistant and 86 were susceptible (Table 2). The data gave good fit to 3 resistant: 1 susceptible ratio ($P = 0.25-0.50$). Among the 98 F₃ families, 52 segregated, 17 were homozygous susceptible and 29 were homozygous resistant (Table 2). The F₃ family segregation was good fit to 7 resistant: 8 segregating : 1 susceptible ($P = 0.10-0.25$).

All F₃ families resistant to pathotype 77-5 were also resistant to pathotype 77-2 but many of the families that were resistant to pathotype 77-2 were susceptible to pathotype 77-5. Therefore, it was concluded that the gene imparting resistance against 77-5 was also operating against 77-2 but the second gene is effective only against pathotype 77-2. Among the designated leaf rust resistance genes (*Lr1-Lr56*), genes *Lr9*, *Lr19*, *Lr24*, *Lr25*, *Lr28*, *Lr29*, *Lr32*, *Lr39*, *Lr41*, *Lr42* and *Lr45* confer resistance to pathotype 77-5 [7&8]. All these genes are also resistant to pathotype 77-2. Three genes namely, *Lr15*, *Lr20*, *Lr26* are resistant to 77-2 but susceptible to 77-5. Therefore, one of the two independent dominant genes of HS424 that imparted resistance to pathotype 77-2 is likely to be either *Lr15* or *Lr20* or *Lr26*, whereas the other gene must be among the genes that are resistant to 77-5 since the

second gene was effective against both pathotypes. The inheritance pattern of HS424 against stem rust pathotypes 40A and 40-1 indicated presence of *Sr24/Lr24* which is effective against pathotype 40A only whereas resistance against both the pathotypes 40A and 40-1 may be ascribed to *Sr31*. Conclusive evidence of the presence of *Lr24* and *Lr26* came from molecular marker study (Fig. 1). A specific 500bp fragment was amplified from the DNA of HS424 and *TcLr24* which confirmed presence of *Sr24/Lr24* in HS424. Similarly, a 1.5kb specific fragment was amplified in HS424 and other lines carrying *Lr26* which confirmed presence of rye chromatin in HS424. Pedigree information substantiated that rye chromatin was due to 1BL.1RS translocation and hence, it was concluded that HS424 also carry *Lr26*. Therefore, the two genes for stem rust resistance in HS424 must be *Sr24* and *Sr31* as these



Fig. 1. Lanes 1-4 : PCR product of *Lr26* marker and lanes 5-7 : PCR product of *Lr24* marker; M: Marker; 1: *TcLr26*; 2: PBW343; 3: HS424; 4: Agra Local; 5: Agra Local; 6: *TcLr24*; 7: HS424

genes are completely linked with *Lr24* and *Lr26*, respectively. HS424 also carries *Yr9* as it is completely linked with *Lr26*. *Yr9* is resistant to all pathotypes of stripe rust except 46S119 and 78S84. Therefore, resistance to pathotype 78S84 must be due to additional seedling resistance gene. It is likely that the additional seedling resistance gene is *Yr2* as it is resistant to pt. 78S84 and prevalent in Indian wheat lines. However, its presence can not be proved conclusively from the present study. Though the seedling of HS424 was susceptible to pt. 46S119 but the adult plant has shown less terminal disease severity (40MS) as compared to the susceptible check Agra Local (100S). Low terminal disease must be due to additional adult plant resistance factors in HS424. A slow rusting stem rust resistance gene, *Sr2*, was also postulated on the basis of microchlorosis, a morphological marker associated with *Sr2* [2&9]. Based on infection type data, inheritance study, morphological marker, genetic linkage and molecular marker analyses it is concluded that HS424 carries genes *L24+Lr26+Sr2+Sr24+Sr31+Yr9*. In addition, it also has adult plant stripe rust resistance gene/s. The rust resistance of varieties released for Northern Hills Zone (NHZ) is mainly dependent on *Lr26+Sr31+Yr9* gene complex [10]. Resistance of HS424 is not solely dependent on 1BL.1RS translocation, hence, it will add additional rust resistance genes in the NHZ.

Acknowledgement

The authors are thankful to the Project Director, DWR, Karnal and Director, NATP for providing facility and financial support, respectively, to carry out studies on wheat rust genetics.

References

1. Nayar S. K., Prashar M. and Bhardwaj S. C. 1997. Manual of current techniques in wheat rusts. Research Bulletin No. 2: p18. Directorate of Wheat Research, Regional Station, Flowerdale, Shimla 171002.
2. Brown G. N. 1997. The inheritance and expression of leaf chlorosis associated with gene *Sr2* for a partial resistance to wheat stem rust. *Euphytica*, **95**: 67-71.
3. Peterson R., Campbell A. B. and Hannah A. E. 1948. A diagrammatic scale for estimating rust severity on leaves and stems of cereals. *Can. J. Res., Sec. C.*, **26**: 496-500.
4. Fransis H. A., Leitch R. A. and Koebner R. M. D. 1995. Conversion of a RAPD generated PCR product, containing a novel repetitive element, in to an fast and robust assay for the presence of rye chromatin in wheat. *Theor. Appl. Genet.*, **90**: 636-642.
5. Mago R., Bariana H. S., Dundas I. S., Spielmeier W., Lawrence G. J., Pryor A. J. and Ellis. 2005. Development of PCR markers for the selection of wheat stem rust resistance genes *Sr24* and *Sr26* in diverse wheat germplasm. *Theor. Appl. Genet.*, **111**: 496-504.
6. Rogers S. O. and Bendich A. J. 1988. Extraction of DNA from plant tissues. In: Gelvin SB and Shilperoot RA (eds.). *Plant molecular biology manual*. Kluwer Academic Publishers, Dordrecht, pp. 1-10.
7. McIntosh R. A., Wellings C. R. and Park R. F. 1995. *Wheat Rusts: An Atlas of Resistance Genes*. CSIRO, Melbourne, Australia.
8. McIntosh R. A., Devos K. M., Dubcovsky J., Rogers W. J., Morris C. F., Appels R. and Anderson O. D. 2006. Catalogue of gene symbols for wheat: 2006 supplement, <http://wheat.pw.usda.gov/ggpages/wgc/2006upd.html>.
9. Nayar S. K., Prashar M., and Bhardwaj S. C. 1999. Characterization of *Lr34* and *Sr2* in Indian wheat (*Triticum aestivum*) germplasm. *Indian J. Agric. Sci.*, **69**: 718-721.
10. Nayar S. K., Nagarajan S., Prashar M., Bhardwaj S. C., Jain S. K. and Datta D. 2001. Revised Catalogue of Genes that Accord Resistance to *Puccinia* species in Wheat. Research Bulletin No.3, 48pp. Directorate of Wheat Research, Regional Station, Flowerdale, Shimla-171002.