Indian J. Genet., 51 (2): 252–259 (1991)

ISOENZYME VARIATION IN CUCUMIS SPP.

V. S. SUJATHA,^{*} V. S. SESHADRI, K. N. SRIVASTAVA AND T. A. MORE

Division of Vegetable Crop and Division of Biochemistry, Indian Agricultural Research Institute, New Delhi 110012

(Received: April 5, 1990; accepted: June 12, 1990)

ABSTRACT

Isoenzyme variation in 13 species of *Cucumis*, including *C. melo* and *C. sativus*, was studied for three enzyme systems, viz. peroxidase, GOT and GDH. The species showed very little similarity for PRX and GDH zymograms. However, there was high degree of similarity among the species for GOT zymogram. All the species possess a genus-specific isozyme at GOT₄. There was very little similarity among the species for the three isozymes put together. No *Cucumis* species of African origin seem to have contributed to the evolution of *C. melo*.

Key words: Isoenzymes, Cucumis spp. percentage similarity.

Isoenzyme variation in 13 species of *Cucumis*, including *C. melo* (x = 12) and *C. sativus* (x = 7), was studied for three enzyme systems to bring out the affinities if any, among the members of the genus *Cucumis*.

1

MATERIALS AND METHODS

The *Cucumis* species compared in this study are listed below along with the origin/source of the materials.

Polyacrylamide gel electrophoresis using vertical slab gel was carried out. Table 1 provides the details of the enzymes assayed.

The loci were represented by a numerical subscript eg. GOT₁, PRX₂, etc. The numbering was done in the ascending order from the fastest to the slowest moving anodal loci.

^{*}Present address: Department of Plantation Crops & Spices, College of Horticulture, Kerala Agricultural University, Vellanikkara 680 654, Kerala. May, 1991]

No. in the figs.	Species/variety	Source			
8	Cucumis melo L.	IARI, New Delhi			
33	C. anguria L. GBNR 1970	Wageningen, Netherland			
34	C. anguria var. longipes (Hook. f.) Meeuse GBNR 1735	Â1			
37	C. africanus L.F. GBNR 1984	"			
38	C. figarei Naud. GBNR 1804	"			
40	C. zeyheri Sond. GBNR 1053 (tetraploid)	"			
41	C. meeusii Jeffrey GBNR 1800	11			
42	C. melo var. agrestis Naud. GBNR 1987	11			
43	C. ficifolius A. Rich GBNR 1801	11			
35	C. myriocarpus Naud. C.73184	South Africa			
36	C. prophetarum L. IC 155	India			
39	C. zeyheri Sond. diploid 73252	South Africa			
45	C. sativus L.	IARI, New Delhi			

MEASUREMENT OF SIMILARITY

The degree of electrophoretic similarity among the *Cucumis* species was calculated by pairwise comparison of the genotypes using the method of Sokal and Sneath [1]. Similarity index (S.I.) was calculated by the following formula.

S.I. = $\frac{\text{No. of homologous bands}}{\text{No. of homologous bands}} + x 100$ nonhomologous bands

The average of the similarity indices for all the enzymes gave average similarity index for pairwise comparisons.

RESULTS

In the peroxidase zymogram, all the taxa with x = 12 possess the fastest moving anodal band PRX₁. This was absent in *Cucumis sativus* (x = 7). At PRX₂ a cluster of bands was found in all the species, but very few isozymes were common among the species. At PRX₃ and PRX₄ also, there was no similarity. On the whole, there was very little similarity among the species for the peroxidase zymogram (Table 2, Fig. 1).

Enzyme	Sample	Gel concentration	Gel buffer	Electrode buffer	Staining technique	
Peroxidase (PRX)	Root & hypocotyl region of 4–5 weeks old seedlings	7% acrylamide	Tris-chloride pH 9.0	T ris -glycine pH 8.3	Adopted from Conklin & Smith [8]	
Esterase (EST)	3-4 days old seedlings	"	*1	17	Shaw & Koen [9]	
Glutamate oxalo acetate transaminase (GOT)	17	9.5% acrylamide	19	"		
Glutamate dehydrogenase (GDH)	"	8% acrylamide	Tris-chloride pH 8.0	**	**	

Table 1. Enzymes assayed in Cucumis species

In the GOT system (Fig. 2), on the other hand, all the species shared a common band at GOT4 under the conditions of GOT analysis. This isozyme was present in *Cucumis sativus* also and absent in other genera like *Praecitrullus* and *Citrullus*. The two *anguria* accessions and *C. prophetarum* had GOT3 isozyme with similar mobility as that of *C. melo*. Apart from *C. prophetarum*, all the wild species of *Cucumis* shared a common band at GOT2. *C. prophetarum* had GOT zymogram similar to that of *C. melo*. The two *anguria* accessions, diploid (73252) and tetraploid (GBNR 1053) *C. zeyheri*, *C. africanus*, and *C. figarei* have an identical band at GOT1. The two *anguria* accessions did not differ in their banding pattern. Thus there was good degree of similarity among the *Cucumis* spp. for the GOT zymogram (Table 2).

In the GDH system (Fig. 3), it was difficult to group the isozymes of different *Cucumis* species under a few loci. The percentage of similarity among the *Cucumis* species for the GDH zymogram is given in Table 2.

Average similarity among the species for the three enzymes is listed in Table 3. As can be seen, none of the species showed appreciable similarity with any other species. The percentage of similarity among the species varied from 8 to 36. *C. melo* and *C. prophetarum* shared 33% common bands between them. The highest S.I. (36%) was between the two *zeyheri* collections. The two *anguria* accessions had 33% S.I. *C. anguria* var. *longipes* had 33% S.I. with *C. melo* also.

C. melo and *C. prophetarum* showed the lowest level of similarity with the remaining species. The two *anguria* accessions showed relatively high level of similarity with all the *Cucumis* species studied.

DISCUSSION

The peroxidase zymogram gave a very complicated picture, having a large number of isozymes with varying mobility in each species (Fig. 1). Even though the banding pattern

May, 1991]

		-				-						
Enzyme	S. I. of different species											
	8	33	34	35	36	37	38	39	40	41	42	43
Peroxidase GOT GDH	100											
Peroxidase GOT GDH	0 50.0 0	100										
Peroxidase GOT GDH	0 50.0 50.0	0 100.0 0	100									
Peroxidase GOT GDH	0 28.6 22.2	0 57.1 0	0 57.1 0	100								
Peroxidase GOT GDH	0 100 0	0 50.0 0	0 50.0 0	0 28.6 0	100							
Peroxidase GOT GDH	11.8 25.0 0	0 75.0 0	0 75.0 0	0 57.1 0	23.5 25.0 0	100						
Peroxidase GOT GDH	0 28.6 0	0 85.7 0	0 85.7 0	0 57.1 0	0 28.6 0	0 85.7 0	100					
Peroxidase GOT GDH	0 28.6 0	0 75.0 0	16.7 75.0 0	0 57.1 0	0 25.0 0	0 75.0 0	0 85.7 0	100				
Peroxidase GOT GDH	0 28.6 0	0 75.0 0	26.7 75.0 0	0 57.0 0	0 25.0 0	0 75.0 0	0 85.7 0	0 75.0 33.3	100 —			
Peroxidase GOT GDH	0 28.6 0	0 57.1 0	13.3 57.1 0	0 66.7 0	0 28.6 0	0 57.1 0	0 66.7 0	0 57.1 0	0 57.1 0	100		
Peroxidase GOT GDH	0 28.6 0	0 57.1 0	0 57.1 0	0 66.7 0	0 28.6 0	0 57.1 0	0 66.7 0	0 57.1 0	0 57.1 0	0 66.7 0	100 —	
Peroxidase GOT GDH	0 28.6 0	0 57.1 0	0 57.1 0	0 66.7 0	0 28.6 0	0 57.1 0	0 66.7 0	0 57.1 0	0 57.1 0	0 66.7 0	0 66.7 0	100

Table 2. Percentage of similarity among Cucumis species for peroxidase, GOT and GDH zymograms

'S. Nos. of species corresponding to those used in figures (cf. Materials and Methods).

was similar, very few isozymes showed identical mobility among the species. Each species had its own individual zymogram. These results are contradictory to Dane's [2] findings, who observed a high degree of similarity among the *Cucumis* spp. for peroxidase zymogram.

V. S. Sujatha et al.

Table 3. Average S.I. among Cucumis species for peroxidase, GOT and GDH

Species	8	33	34	35	36	37	38	39	40	41	42	43
8	100											
33	17.0	100										
34	33.0	33.0	100									
35	17.0	19.0	19.0	100								
36	33.0	17.0	17.0	10.0	100							
37	12.25	25.0	25.0	19.0	16.0	100			<i>,</i>			
38	10.0	29.0	29.0	19.0	10.0	29.0	100					
39	8.0	25.0	31.0	19.0	8.0	25.0	29.0	100				
40	8.0	25.0	34.0	19.0	8.0	25.0	29 .0	36.0	100			
41	10.0	19.0	23.0	22.0	10.0	19.0	22.0	19.0	19.0	100		
42	10.0	19.0	1 9 .0	22.0	10.0	19.0	22.0	19.0	19.0	22.0	100	
43	10.0	19.0	19.0	22.0	10.0	19.0	22.0	19.0	19.0	22.0	22.0	100

^{*}Cf. note to Table 2.

On the other hand, Esquinas-Alcazar [3] reported seven different clusters in the species he studied. The two different clusters observed in the cross-compatible species were found to be different from *C. melo.* Similarly, different species showed more differences than similarity in the GDH zymogram also.

In respect of GOT, all the *Cucumis* species studied showed an identical band at GOT4 (Fig. 2). This band was present in *C. sativus* also, but was absent in other genera like *Citrullus* and *Praecitrullus*. Thus, this isozyme could be a characteristic of the genus *Cucumis*. While all the African wild species (x = 12) showed an identical band at GOT₂, the Indian collection of *C. prophetarum* (x = 12) had an isozyme similar to *C. melo*. The two *anguria* accessions studied had an isozyme at GOT₂ with similar mobility to that of *C. melo* and *C. prophetarum*. The *anguria* collections, diploid and tetraploid *C. zeyheri*, *C. figarei* and *C. africanus* had a similar isozyme at GOT₁. *C. prophetarum* had an identical GOT zymogram with *C. melo*. The two *anguria* accessions also had similar banding pattern. These results are in agreement with the findings of Dane and Esquinas-Alcazar [3] who also found high degree of similarity among the species for GOT.

Summing up the results on isozyme variation in *Cucumis* species, we can conclude that the different species possess their own individual zymogram. There was very little similarity among the species for the three enzyme systems put together. However, there was some similarity among the species, including *C. sativus*, especially for the GOT zymogram, which confirms that all these taxa belong to the same genus, *Cucumis*.

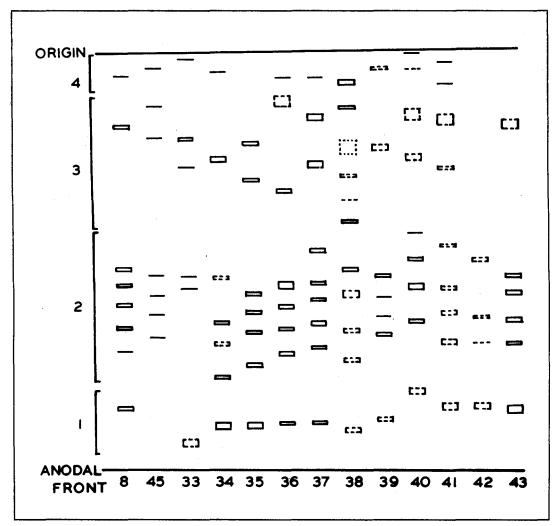


Fig. 1. Peroxidase isozyme pattern of Cucumis spp.

On an average, the two *anguria* accessions, *C. anguria* and *C. anguria* var. *longipes*, showed more similarity with other species than any other of the remaining species. On the other hand, *C. melo* and *C. prophetarum* shared minimum similarity with other species. However, they had high S.I. (33%) between them.

C. prophetarum from India is reported to have high degree of cytological homology with C. melo [4]. Dane [2] concluded that C. prophetarum from India may probably be C. melo and appears to be misclassified. In the present study, although C. prophetarum has shown an

V. S. Sujatha et al.

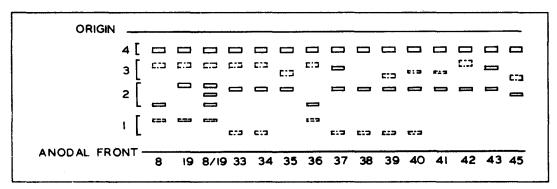


Fig. 2. Variation in GOT zymogram of Cucumis spp.

identical GOT zymogram with *C. melo*, the peroxidase and GDH zymograms of the two species were different, thus, justifying their inclusion under separate taxa.

The results of the present study are contradictory to the findings of Dane [2], who found a large degree of similarity among the *Cucumis* species for the peroxidase zymogram. Our results also do not agree with those of Esquinas-Alcazar [3], who classified *Cucumis* species into four groups based on Nei's genetic distance between them.

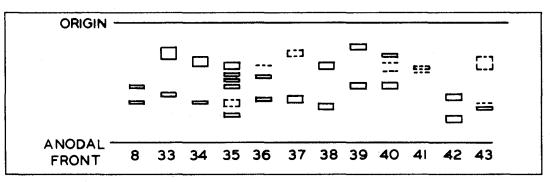


Fig. 3. Variation of GDH isozymes in Cucumis spp.

These observations lend support to the view that no *Cucumis* species (with x = 12) of African origin contributed to, or is involved in, the evolution of *C. melo* or the common muskmelon. Isozyme studies do corroborate the observations of a certain degree of remoteness of *C. melo* from the other 12-chromosome species found in the crossability studies [5–7]. In other words, the diversification within *C. melo* and apparent morphological variation within the taxon had no relationship within *C. melo* and apparent large morphological variation within the taxon had no relationship with the other 12-chromosome species of African origin.

ACKNOWLEDGEMENTS

The authors are grateful to Dr. K. R. Sarkar, Division of Genetics, IARI, New Delhi and Dr. I. M. Santha, Division of Biochemistry, IARI, for their valuable suggestions during the course of work. The financial assistance provided to the first author by the Indian Agricultural Research Institute, New Delhi, in the form of a Senior Research Fellowship is gratefully acknowledged. The results reported in this communication formed part of the thesis for the Ph.D. degree of the Indian Agricultural Research Institute by the first author.

REFERENCES

- 1. R. R. Sokal and P. H. A. Sneath. 1963. Principles of Numerical Taxonomy. N. H. Freeman and Company, San Francisco.
- F. Dane. 1983. Cucurbits. In: Isozymes in Plant Genetics and Breeding. Part B (eds. S. D. Tanksley and T. J. Orton). Elsevier Science Publishers, B. V. Amsterdam: 369–382.
- 3. J. T. Esquinas-Alcazar. 1981. Alloenzyme variation and relationship among Spanish land races of *Cucumis melo* L. Kulturpilanze, 22:337–352.
- 4. C. Ramachandran. 1982. Cytogenetical Studies in *Cucumis*. Ph.D. Thesis, Indian Agricultural Research Institute, New Delhi: 127.
- 5. J. R. Deakin, G. W. Bohn and T. W. Whitaker. 1971. Interspecific hybridization in *Cucumis*. Econ. Bot., 25: 195-211.
- 6. R. W. Robinson and E. Kowalewski. 1978. Interspecific hybridization of *Cucumis*. Cucurbit Genetics Cooperative, **1**: 40.
- 7. F. Dane, D. W. Denna and T. Tsuchiya. 1980. Evolutionary studies of wild species in the genus *Cucumis*. Z. Pflanzenzuchtg., 85: 89–109.
- 8. M. E. Conklin and H. H. Smith. 1971. Peroxidase isoenzymes. A measure of molecular variation in ten species of *Datura*. American J. Bot., 58(7): 688–696.
- 9. C. R. Shaw and A. L. Koen. 1968. Starch gel zone electrophoretics. *In*: Chromatographic and Electrophoretic Techniques, vol. 2 (ed. I. Smith). John Wiley, New York.

۴.

٤