

# **Sequence diversification and phylogenetic analysis of selfincompatibility specific determinant genes in Brassicaceae**

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#### **Abstract**

**It is a long history of theoretical interest in understanding the genetic causes and consequences of shifts between outcrossing and inbreeding in plant mating systems. Selfincompatibility (SI) systems genetically promote outcrossing, and its transitions to inbreeding potentially changes genetic diversity in the evolution of species. SI in the Brassicaceae family is sporophytically controlled by a single locus (S-locus), which locates the female determinant gene SRK (S-locus receptor kinase) and the male determinant gene SCR (S-locus cysteine rich). Operation of diversifying selection maintains the diversity of SRK and SCR during the evolutionary course of S haplotypes. Here we used deduced amino acid sequences of SRK and/ or SCR publicly available from seven species in three genera, Brassica, Arabidopsis and Raphanus, to investigate the sequence diversification and phylogenetic distribution of S haplotypes in the Brassicaceae family. The results reveal that SCR shows higher sequence distance than SRK within or between genera, and also in the interspecific pairs of S haplotypes. Phylogenetic analysis supports the conclusion that SI has a single origin but evolves differently within the Brassicaceae. The genetic mechanisms and consequences of the loss of SI in the Brassicaceae family are discussed.**

**Key words**: Self-incompatibility, SRK, SCR, phylogenetic analysis, Brassicaceae

#### **Introduction**

Many flowering plants have a genetically controlled mechanism such as self-incompatibility (SI) to avoid potentially negative consequences of inbreeding. SI has been extensively studied in the Brassicaceae family and is sporophytically controlled by a single major locus, S-locus (Bateman 1955). The S-locus is comprised of a number of completely linked genes and segregates as an S haplotype (Nasrallah and

Nasrallah 1993). The female determinant SRK (S-locus receptor kinase) (Takasaki et al. 2000) and the male determinant, SCR/SP11 (S-locus cysteine-rich protein/ S-locus protein 11) (Schopfer et al. 1999; Suzuki et al. 1999), are involved in the SI recognition reaction in Brassicaceae. Many species in Brassicaceae are important resources for SI studies including self-fertile model plant Arabidopsis thaliana and its relatives, selfincompatible A. lyrata, and major vegetables and crops in Brassica and Raphanus genera. Breakdown of SI is an important topic which has been greatly studied in Brassicaceae. Two mechanisms have been proposed to account for the loss of SI. The major one is mutations of SRK or SCR, like A. thaliana (Tsuchimatsu et al. 2010) and B. rapa (Fujimoto et al. 2006). Another is the interruption of downstream components of the signal transduction pathway like B. oleracea (Cabrillac et al. 2001).

Due to the strong negative frequency-dependent selection, the SI system typically maintains a large number of haplotypes (Lawrence 2000). The distribution of S haplotypes has been well analyzed in some genera, in which some characteristic differences were found. For example, the Brassica S haplotypes can be divided into two dominant classes, class I and class II. Class I S haplotypes are dominant over class II S haplotypes in the pollen side but co-dominant in the stigma side (Nasrallah et al. 1991). Most of the previous phylogenetic analyses used either two genera or incomplete DNA sequences to dissect the evolution and genetic diversity of S haplotypes (Sato et al. 2002). But there are scanty reports using the protein data from both wild species and cultivated species for

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phylogenetic analysis, in view that mature peptide region is better to discuss on the evolutionary point of SI determinant genes. In our previous study, an S haplotype was characterized from a self-compatible B. rapa cultivar Dahuangyoucai, which was proved to highly similar to S-f2 haplotype found in another selfcompatible B. rapa Yellow sarson, which gave rise to the confusion on the origin of S haplotype and genetic evolution (Zhang et al. 2013).

In the present study, phylogenetic analysis was performed for the first time with mature protein sequences of SRK and SCR alleles publicly available from seven Brassicaceae species belonging to three genera. The results will be helpful to understand the genetic diversity and evolution of S haplotypes in Brassicaceae and dissect the genetic causes and consequences of the breakdown of SI in higher plants.

#### **Materials and methods**

## **Sequences collecting**

Sequences of S-locus genes were collected using genus name and gene name as keywords in Genbank of NCBI website (www.ncbi.nlm.nih.gov). Among all the searched sequences, those with complete CDS were chosen and their amino acid sequences were then collected. The sequence was named based on the species and S haplotype, for example, SRK of S8 in B. rapa was described as BraSRK8 but for A. thaliana, the S haplotype was not denominated, so the ecotype was used as suffix, like AthSRK-Kas2 for SRK of A. thaliana ecotype Kas-2.

#### **Phylogenetic analysis**

All sequences were multiple aligned using CLUSTAL W program (Thompson et al. 1994) following the default parameters: for pairwise alignments, a gap opening penalty=10 and a gap extend penalty =0.1; for multiple alignments, a gap opening penalty=10 and a gap extend penalty=0.2. The delay divergent sequences  $= 30\%$ , DNA transition weight  $=0.5$  and use negative matrix OFF. Nucleotide sequence distance between and within each genus was calculated using MEGA3.0 (Kumar et al. 2004) following the parameters, gap/ missing data: pairwise deletion; model: nucleotide, p distance; pattern among lineages: same (homogeneous); rates among sites: uniform rates. Phylogenetic trees were constructed with deduced amino acid sequences using the minimum-evolution method of MEGA3.0. The default parameters included gap/missing data, pairwise deletion; model, amino: poisson correction; substitutions to include, All; cut-

off value for consensus/condensed tree, 50%. The bootstrap probabilities of 1000 trials were used for phylogeny test. No sequences appropriate for rooting the tree were found, so the tree was left unrooted.

#### **Results**

#### **Sequence collection of SI determinant genes**

A total of 35 SCR and 31 SRK from seven species of three genera were searched. The sequences included the predominant model plant, A. thaliana; the major Brassica crop, B. napus and the major vegetables, B. rapa, B. oleracea and R. sativus. These species showed different S phenotype intra-genus, like selfincompatible A. lyrata and self-compatible A. thaliana and also included the cultivated Brassicaceae species B. rapa, B. napus and the wild species A. halleri.

The length of SRK changed from 1,461 bp to 2,583 bp, and the amino acid length was from 795 to 860. BraSRK733 has eight exons and BraSRK734 has nine exons, which both were obviously different from other SRK genes with seven exons. The length of SCR was varied from 225 bp to 288 bp, and the corresponding protein was from 74 to 95 amino acids. In Brassica, all the class I SCRs were shorter than that of class II in nucleotide length. Inter-specific S haplotype pair showed highly similar SRK and SCR, like BolS-15, BraS-60 and BnaS-6. They showed 98% (two bp difference of the 288 bp) and 99% (two bp difference of the 2571 bp) nucleotide sequence identity for the SCR and SRK, respectively.

#### **Sequence diversification of SI determinant genes**

Nucleotide sequence distance between and within the three genera were calculated (Table 1). SCR genes showed higher degree of sequence distance within Brassica (0.43) than other two genera, Raphanus (0.20) and Arabidopsis (0.32). It exhibited nearly the same level of polymorphism between genera, Brassica and

**Table 1.** The nucleotide sequences distance between and within the genus

Gene	Genus	Bra- ssica	nus	dopsis	Rapha- Arabi- Overall
<b>SCR</b>	Brassica Raphanus Arabidopsis	0.43 $0.46*$ $0.48*$	0.20 $0.60*$	0.32	0.44
<b>SRK</b>	Brassica Raphanus Arabidopsis	0.15 $0.14*$ $0.25*$	$0.25*$	በ 17	0.20

\*,distance between genus

Raphanus (0.46), Brassica and Arabidopsis (0.48), but both were lower than that between Raphanus and Arabidopsis (0.60). SRK sequences displayed a moderate polymorphism i.e., the mean sequence distance within each group is 0.15 for Brassica and 0.17 for Arabidopsis. In view of only one SRK of Raphanus collected, it was impossible for calculating the distance within Raphanus. The distance within Brassica (0.15) was comparable to that between it and Raphanus (0.14), while both of them showed the same distance from Arabidopsis (0.25). For the overall mean distance of the nucleotide sequence, SCR displayed striking diversification (0.44) than SRK (0.20). Sequence distances of two class S haplotypes in Brassica were also calculated (Table 2). Class I showed lower sequence distance than class II, like SRK within class I and class II were 0.13 and 0.03, respectively, while SCR within class I was 0.33 and within class II

Table 2. The nucleotide sequences distance of S haplotypes in Brassica

Gene	Class			
<b>SCR</b>		0.33 $0.12*$	0.59	
<b>SRK</b>		0.13 $0.21*$	0.03	

\*,distance between class

was 0.59. Besides above mentioned sequence distances, SRK manifested high polymorphism between classes than SCR (distance 0.12 for SCR and 0.21 for SRK) in view of the length difference.

## **Phylogenetic trees constructed using SRK sequences**

The interspecific pair S haplotypes were clustered together with 100% bootstrap support, such as BolSRK12 and BnaSRK1, BraSRK54 and BolSRK60, reflecting the rationality of the phylogenetic tree of SRK (Fig. 1). All the S haplotypes from Brassica including class I Brassica S haplotypes, class II S haplotypes and one RsaSRKa formed a major group, except two Brassica S haplotypes, BraSRK733 and BraSRK734, with longer branches than other haplotypes were clustered with Arabidopsis S haplotype clade. AlySRK6 and AlySRKb were seperated as a deviant clade.

# **Phylogenetic trees constructed using SCR sequences**

In the SCR tree constructed using 35 SCR protein





sequences (Fig. 2), class I and class II S haplotypes of Brassica did not form a monophyletic group, which was obvious in the SRK tree. The same gene sequenced by different researchers, BoISCR13 and BolSCR13b, and the interspecific pair consisting of one haplotype from each of B. oleracea (BolSCR12) and B. rapa (BraSCR47), were particularly clustered with 100% bootstrap value. Most of the class I haplotypes in Brassica were sorted to one clade, except that five S haplotypes were more closely related to Brassica class II S haplotypes and five Raphanus S haplotypes. All the class II S haplotype from Brassica and five S haplotype from Raphanus were assigned into the same group, indicating that the five S haplotypes in Raphanus might belong to class II. All Arabidopsis SCRs was formed into one group, including AlySCR6 and AlySCRb, even without wellsupported bootstrap values, which was different from the profile of the SRK tree.

# **Genetic evolution of SI determinant genes in Brassicaceae**

Combining the two phylogenetic trees, we found the general evolution model in Brassicaceae. Brassica/ Raphanus split from Arabidopsis first, following by the



**Fig. 2. Minimum-evolution tree constructed with the deduced amino acid of SCR: Brassica class I SCR (triangle); Brassica class II SCR (square). One thousand bootstrap trials were performed and the bootstrap values are shown beside the branch**

divergence of S haplotypes before the species differentiation within genus. In Brassica, class I and class II S haplotypes diverge predate the species differentiation, then S alleles within class I diversify more ancient than those within class II.

#### **Discussion**

The situation that fewer recessive haplotypes at high frequency and a large number of dominant haplotypes at lower frequencies was predicted in Brassica (Billiard et al. 2007). It was also reflected in the present study. Greater sequence distance was observed in SCR alleles than in SRK alleles, but the phylogenetic analyses showed the similar topology, suggesting a positive correlation in divergence time between the female and male determinants. A variety of factors resulted in the high sequence diversification of S haplotypes. Little or no recombination occurs between SRK and SCR promotes the maintenance of extensive nucleotide diversity (Charlesworth et al. 2003). Gene

conversion between SRK and SLG (S-locus glycoprotein) also plays a significant role (Sato et al. 2002). Wild and cultivated Brassicaceae species share similar levels of SRK diversity, indicating that domestication has had but a minor effect on S-locus diversity (Edh et al. 2009).

In order to highlight the pending questions, great progresses have been made in Brassicaceae (Tedder et al. 2011). Genetic and phylogenetic features of SI in the tribe Biscutelleae were observed departing from patterns in the reference Arabidopsis, while similar to Brassica (Leducq et al. 2014). Although the phylogenetic trees constructed using two SI determinants were unrooted, they showed similar pattern that the interspecific S haplotype pairs were closely clustered. Phylogenetic results supported the previous conclusion that SI has a single origin but evolves differently in the Brassicaceae family. Brassica/Raphanus split from Arabidopsis first, predating the divergence of S haplotypes followed by the species differentiation within genus (Zhang et al. 2011). Five class I S haplotypes were closely clustered to the major lineage of class II in the SCR tree, might be due to the synonymous and non-synonymous amino acid substitution rate (Tedder et al. 2011). Previous results found a single polymorphic 24 nucleotide small RNA, named SP11 methylation inducer 2 (Smi2), controls the linear dominance hierarchy of the four class II SP11 alleles in B. rapa (Yasuda et al. 2016). Multilayered dominance hierarchy was put forward to understand the dominance regulatory mechanism of SI in the Brassicaceae (Fujii and Takayama 2018).

As a longstanding subject, the genetic causes and consequences of loss of SI have received theoretical and empirical attention since the time of Darwin. In Brassicaceae, the model plant A. thaliana and cultivated plants in the genus Brassica, together with the genus Capsella have been extensively studied. Mutation of S-locus was generally thought to be the main causes for the breakdown of SI (Rea et al. 2010). Comparisons between multiple populations in selfing and outcrossing species in the Brassicaceae were helpful to evaluate the consequences of mating system differences. Further investigation should combining the sequence data of emerging genomic information, population structure, changes in effective population size and genetic diversity from cultivated plants to wild populations in relation to mating system, together with the individual variations such as maternal effects, background genetics, the history of outcrossing and possibly particular S haplotypes.

## **Authors' contribution**

Conceptualization of research (XZ); Designing of the experiments (XZ, YL); Contribution of experimental materials (YL); Execution of field/lab experiments and data collection (XM, WZ); Analysis of data and interpretation (XM, WZ); Preparation of manuscript  $(XZ)$ .

## **Declaration**

The authors declare no conflict of interest.

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## **References**

- Bateman A. J. 1955. Self-incompatibility systems in angiosperms.III. Cruciferae, Heredity, **9**: 53-68.
- Billiard S., Castric V. and Vekemans X. 2007. A general model to explore complex dominance patterns in plant sporophytic self-incompatibility systems. Genetics, **175**: 1351-1369.
- Cabrillac D., Cock J. M., Dumas C. and Gaude T. 2001. The S-locus receptor kinase is inhibited by thioredoxins and activated by pollen coat proteins. Nature, **410**: 220-223.
- Charlesworth D., Bartolomé C., Schierup M. H. and Mable B. K. 2003. Haplotype structure of the stigmatic selfincompatibility gene in natural populations of Arabidopsis lyrata. Mol. Biol. Evol., **20**: 1741-1753.
- Edh K., Widén B. and Ceplitis A. 2009. The evolution and diversification of S-locus haplotypes in the Brassicaceae family. Genetics, **181**: 977-984.
- Fujii S and Takayama S. 2018. Multilayered dominance hierarchy in plant self-incompatibility. Plant Reprod. **31**(1):15-19.
- Fujimoto R., Sugimura T. and Nishio T. 2006. Gene conversion from SLG to SRK resulting in selfcompatibility in Brassica rapa. FEBS Letters, **580**(2): 425-430.
- Kumar S., Tamura K. and Nei M. 2004. MEGA3: integrated software for molecular evolutionary genetics analysis and sequence alignment. Bioinformatics, **5**: 150-163.
- Lawrence M. J. 2000. Population genetics of the homomorphic self-incompatibility polymorphisms in flowering plants. Ann. Bot., **85**: 221-226.
- Leducq J-B, Gosset C-C, Calin K, Schmitt É, Castric V and Vekemans X. 2014. Self-Incompatibility in Brassicaceae: Identification and Characterization of SRK-Like Sequences Linked to the S-Locus in the Tribe Biscutelleae. Genes Genomes Genetics, **4**: 983-992.
- Nasrallah J. B. and Nasrallah M. E. 1993. Pollen-stigma signaling in the sporophytic self-incompatibility

response. Plant Cell, **5**: 1325-133.

- Nasrallah J. B., Nishio T. and Nasrallah M. E. 1991. The self-incompatibility genes of Brassica: expression and use in genetic ablation of floral tissues. Ann. Rev. Plant Physiol. Plant Mol. Biol., **42**: 393-422.
- Rea A. C., Liu P. and Nasrallah J. B. 2010. A transgenic self-incompatible Arabidopsis thaliana model for evolutionary and mechanistic studies crucifer selfincompatibility. J. Exp. Bot., **61**(7): 1897-1906.
- Sato K., Nishio T., Kimura R., Kusaba M., Suzuki T., Hatakeyama K., Ockendon D. and Satta Y. 2002. Co-evolution of the S-locus genes, SRK, SLG and SP11/SCR, in Brassica oleracea and Brassica rapa. Genetics, **162**: 931-940.
- Schopfer C. R., Nasrallah M. E. and Nasrallah J. B. 1999. The male determinant of self-incompatibility in Brassica. Science, **286**: 1697-1700.
- Suzuki G., Kai N., Hirose T., Fukai K., Nishio T., Takayama S., Isogai A., Watanabe M. and Hinata K. 1999. Genomic organization of the S locus: Identification and characterization of genes in SLG/SRK region of S(9) haplotype of Brassica campestris (syn. rapa). Genetics, **153**(1): 391-400.
- Takasaki T., Hatakeyama K., Suzuki G., Watanabe M., Isogai A. and Hinata K. 2000. The S receptor kinase determines self-incompatibility in Brassica stigma. Nature, **403**: 913-916.
- Tedder A., Ansell S. W., Lao X., Vogel J. C. and Mable B. K. 2011. Sporophytic self-incompatibility genes and mating system variation in Arabis alpine. Ann. Bot., **157**: 1-15.
- Thompson J. D., Higgins D. G. and Gibson T. J. 1994. CLUSTAL W, improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. Nucleic Acids Res., **22**(22): 4673- 4680.
- Tsuchimatsu T., Suwabe K., Shimizu-Inatsugi R., Isokawa S., Pavlidis P., Städler T., Suzuki G., Takayama S., Watanabe M. and Shimizu K. K. 2010. Evolution of self-compatibility in Arabidopsis by a mutation in the male specificity gene. Nature, **464**: 1342-1346.
- Yasuda S., Wada Y., Kakizaki T., Tarutani Y., Miura-Uno E., Murase KFujii S., Hioki T., Shimoda T., Takada Y., Shiba H., Takasaki-Yasuda T., Suzuki G., Watanabe M. and Takayama S. 2016. A complex dominance hierarchy is controlled by polymorphism of small RNAs and their targets. Nat. Plants, **3**: 16206.
- Zhang X. G., Yin D. M., Ma C. Z. and Fu T. D. 2011. Phylogenetic analysis of S-locus genes reveals the complicated evolution relationship of S haplotypes in Brassica. Plant Mol. Biol. Rep., **29**: 481-488.
- Zhang X. G., Ma C. Z., Yin D. M., Zhu W., Gao C. B. and Fu T. D. 2013. Characterization of S haplotype in a new self-compatible Brassica rapa cultivar Dahuangyoucai. Czech J. Genet. Plant Breed., **49**(4): 157-163.